

IMPACT OF THERMAL STRESS AND HIGH VPD ON GAS EXCHANGE AND CHLOROPHYLL FLUORESCENCE OF *CITRUS GRANDIS* UNDER DESERT CONDITIONS

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Abstract

The photosynthetic response of *Citrus grandis* to high light intensities, low air humidity and high temperature stress was investigated under desert conditions in the southern Arava Valley (Israel). During summer, a typical midday stomatal closure was observed even in well-watered trees due to the dramatic increase of the leaf-to-air water vapour deficit. As a result of the reduced transpirational cooling, leaf temperatures increased up to 11 °C above ambient air temperature. The combination of heat stress and photoinhibition resulted in a reversible decrease of photosynthetic activity of *Citrus grandis* under the extreme summer conditions.

1. Introduction

Citrus, native of subtropical and tropical regions in Eastern Asia became widely cultivated in areas with Mediterranean climate. In Israel, *Citrus grandis*, *C. sinensis*, *C. limon* and *C. paradisi* are major fruit crops of the coastal plain, where the mean annual rainfall ranges between 500 and 700 mm (Bielorari *et al.*, 1973) and average temperature at midday in the August is 30 °C (Cohen *et al.*, 1997).

During recent years cultivation of *Citrus* has substantially increased under desert conditions in the Negev and in the Arava Valley of Israel. In these arid areas crop production requires intensive irrigation. The climate of the Arava is characterized by mild winters and very hot summers with high air temperatures (> 42 °C), low air humidity and high radiation. Optimum temperature for *Citrus* photosynthesis varies from 25°C to 30°C, while temperatures above 35°C reduce photosynthetic activity (Spiegel & Goldschmidt, 1996). High temperatures combined with low air humidity may induce stomatal closure. In addition the combination of high irradiation and stress conditions, which limit photosynthetic energy conversion by reducing CO₂ supply, may result in enhanced photoinhibition (Baker, 1993; Blanke, 2000; Herppich 2000). Photosystem II is that component of the photosynthetic apparatus which is most sensitive to heat (Bilger *et al.*, 1987) and high light stress (Baker, 1993). In the present study, the response of transpiration, stomatal opening, CO₂ uptake and chlorophyll fluorescence of *Citrus grandis* was investigated under extreme desert conditions.

2. Materials and methods

Three year old *Citrus grandis* trees were investigated during June and December 1997 at the Arava Research Station in Yotvata, Israel (29° 53' N, 33° 3' E), 40 km north of Eilat. The orchard was irrigated automatically by a drip system every second day during summer and every fourth day during winter. The soil was an Arava loamy sand.

Net CO₂ exchange and transpiration of fully expanded leaves were measured with a portable CO₂/H₂O porometer system (HCM-1000, Walz GmbH, Effeltrich, Germany) under ambient conditions according to Veste & Herppich (1995) and von Willert *et al.* (1995). Ambient CO₂ concentration was nearly constant at 350 ppm throughout the day. Area (projected leaf area) based gas exchange parameters were calculated after von Caemmerer and Farquhar (1981). Simultaneously with gas exchange, chlorophyll fluorescence was monitored on the same plant with a Mini-PAM fluorometer (H. Walz GmbH, Effeltrich, Germany). Maximum photochemical efficiency of PSII ($F_v/F_m = F_m - F_0 / F_m$) was determined on dark-adapted (10 to 12 min) attached leaves (n = 7). F_m , F_v and F_0 denote the maximum, the variable and the initial fluorescence, respectively (cf. von Willert *et al.* 1995). Quantum yield of linear electron transport ($F'_m - F / F'_m = \Phi_{PSII}$) were analysed as summarized by von Willert *et al.* (1995) and electron transport rate (J_e) was calculated according to Krall & Edwards (1992) as $J_e = \Phi_{PSII} * 0.5 * 0.84 * PPF$ (PPFD = photosynthetic photon flux density).

3. Results

Typical diurnal time courses of net CO₂ exchange (J_{CO_2}), transpiration (J_{H_2O}), leaf conductance (g_{H_2O}) of *Citrus grandis* and the microclimatic conditions on a normal summer and winter day are shown in Fig. 1. Maximum net CO₂ exchange rates of *C. grandis* typically ranged between 8 and 12 $\mu\text{mol m}^{-2} \text{s}^{-1}$ throughout the year. Both photosynthetic electron transport (J_e , Fig. 2) and net CO₂ exchange (data not shown) were always light saturated at PPFD above 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$. During winter month diurnal courses of leaf conductance, net CO₂ exchange and transpiration were very similar, and no midday depression of gas exchange occurred (Fig. 1). Maximum air temperatures on cloudless winter days reached 20 to 22 °C and mean maximum Δw did not exceed 20 kPa MPa⁻¹.

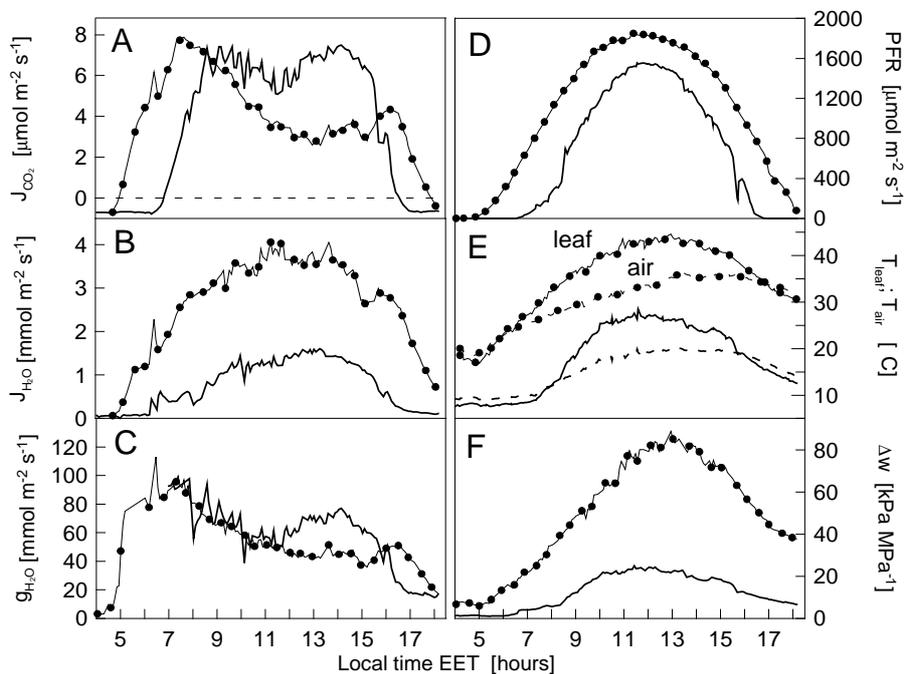


Figure 1: Typical diurnal courses of net CO₂ exchange (J_{CO_2} , A), transpiration (J_{H_2O} , B), leaf conductance (g_{H_2O} , C), photon flux density (PPFD, D), leaf and air temperature (T_{air} , T_{leaf} , E) and air-to-leaf water vapour deficit (Δw , F) in June (circles) and December (solid line).

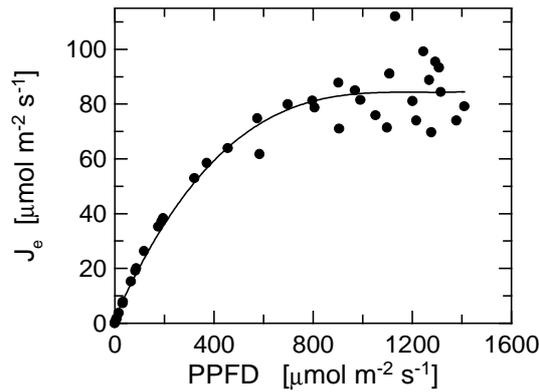


Figure 2: Light response of photosynthetic electron transport (J_e) of *Citrus grandis* in winter.

In contrast, mean maximum air temperatures varied from 33 to 37 °C during summer days. The temperature of sun-exposed leaves raised up to 11 °C above air temperature during hours of high radiation. As a result, Δw increased to more than 85 kPa MPa⁻¹ (Fig. 1). Due to the harsh substantially decreased after ca 9:00 even in well-watered plants.

Midday stomatal closure reduced net CO₂ uptake by nearly 65%. Additionally, high temperature in combination with high irradiance directly affected photosynthesis (Fig. 4). This was indicated by the increase of internal CO₂ concentration around noon (Fig. 4B). Furthermore, maximum photochemical efficiency (F_v/F_m) decreased by 12% from pre-dawn to noon (Fig. 4C). The initial fluorescence (F_0) was nearly unaffected in the early morning hours and tended to rise slightly when leaf temperature exceeded 27°C. At leaf temperatures of 43°C the initial fluorescence was 15% higher than pre-dawn levels (Fig. 4C).

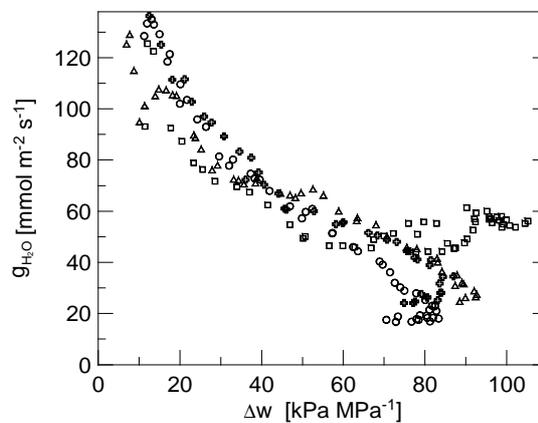


Figure 3: Plot of leaf conductance for water vapour over air-to-leaf water vapour deficit. Given are the results of four summer days (June).

Air temperatures occasionally exceeded 40 °C. On these occasions leaf temperatures of sun-exposed leaves increased up to 47.5 °C. Under these conditions, J_{CO_2} was seriously reduced by the pronounced stomatal closure. Interestingly, leaf conductance at maximum Δw (max. $T_{leaf} = 47.6$ °C) was higher on that hot day than on cooler days (max. $T_{leaf} < 43$ °C). Maximum transpiration rate increased from 3.57 to 5.9 mmol m⁻² s⁻¹.

This higher transpiration rate significantly contributed to the cooling of the leaf and might have prevented the increase of leaf temperature above a critical point. Heat stress above leaf temperatures of 45°C lead to a decrease in the photosynthetic activity (Fig. 4). Nevertheless, in these heat stressed leaves midday internal CO₂ concentration (c_i) was nearly 50 ppm higher ($c_i = 253$ ppm) as compared to leaves experiencing normal summer temperatures (Fig. 4B) and F_v/F_m declined to a lower level (Fig. 4D). The mean initial fluorescence rose by 25% from 436 to 537 when leaf temperatures increased from 45°C to 47.6 °C indicating serious heat stress (Fig 4C). Both F_0 and F_v/F_m recovered during the afternoon when leaf temperature and radiation declined again (Fig. 4D).

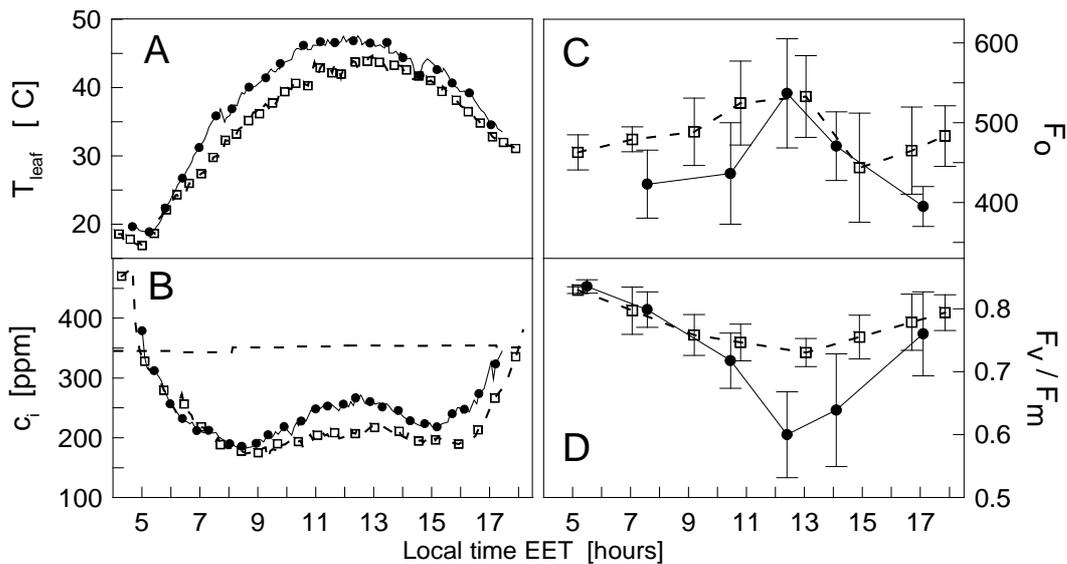


Figure 4: Diurnal courses of leaf temperature (T_{leaf} , A), internal CO₂ concentration (c_i , B), initial fluorescence (F_0 , C) and maximum quantum yield of PS II (F_v/F_m , D) on a normal summer day (squares) and under heat stress (circles).

4. Discussion

Maximum net CO₂ exchange rates of *Citrus grandis* observed in this study under extreme desert conditions corresponded well with values (4 to 12 $\mu\text{mol m}^{-2} \text{s}^{-1}$) obtained for *Citrus sinensis* and *Citrus paradisi* under moderate environmental conditions (Singlair & Allen, 1982; Spiegel & Goldschmidt, 1996; Blanke, 2000). Photosynthesis of citrus was saturated at relatively low light intensities around 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$ in this study. Low light saturation between 600 to 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$ are well known for other *Citrus* species (Singlair & Allen, 1982; Syvertsen, 1984; Braake & Allen, 1995). This indicates that sun-exposed leaves are exposed to an excess of light during most of sunny cloudless days. This easily explains why maximum CO₂ exchange rates did not differ between summer and winter as PPFD were always above the point of light saturation.

A negative influence of Δw on the stomatal opening was observed also in this study for well-irrigated citrus trees in the dry summertime as found for other *Citrus* cultivars (Hall *et al.*, 1975; Kriedemann & Baars, 1981; Braake & Allen, 1995). A midday depression of stomata opening is typical for other well-irrigated fruit trees such as apricots (*Prunus armeniaca*) and grapes (*Vitis vinifera*) (Lange & Meyer, 1979). Syvertsen (1982) hypothesized that the direct effect of Δw on leaf conductance may explain similar transpiration rates under different environmental conditions. However in

our case, leaf conductance in *Citrus grandis* was similar in winter and summer and transpiration rates were even higher in summer. Thus, it may be assumed that *Citrus* has to reduce the maximum transpiration rates because water uptake and transportation capacity are limited by a low hydraulic conductance (Kriedemann & Baars, 1981; Moreschet *et al.*, 1990). The fact that the midday stomatal closure in the fruit trees is controlled by Δw has an important implication on the irrigation management. Increasing the soil water content by applying a greater irrigation volume will not result in higher leaf conductance and higher CO_2 uptake midday (Lange & Meyer, 1979). As a result of the low transpiration rates leaf cooling was reduced and leaf temperatures exceeded air temperatures up to 11 °C. This overheating of the leaves caused a drastic increase of Δw , thus negatively affecting leaf conductance and CO_2 uptake. If CO_2 uptake was only restricted by stomatal closure than the internal CO_2 concentration should decrease. However, under those high temperatures observed in our study c_i significantly increased. This clearly indicates photoinhibition and/or other down-regulating systems (Clifford *et al.*, 1997; Herppich *et al.*, 1997). High photon flux density may induce photoinhibition by overexcitation of photosystem II when stomatal closure limited CO_2 supply (Baker 1993). Furthermore, photorespiration may increase under high light intensities and high temperatures thus contributing to the increase of c_i . In the investigation presented, PSII was not much affected in comparison to the investigations of Werner *et al.* (1999) and Herppich (2000). F_v/F_m declined only by 12% even at leaf temperatures above 42 °C. In addition, predawn values of F_v/F_m were relatively high if compared with other evergreen Mediterranean trees (Werner *et al.*, 1999). Photorespiration provides an effective electron sink and may help prevent excessive reduction of electron transport chain and photoinactivation of electron transport during light stress (Heber *et al.*, 1996).

An increase of photorespiration may explain the low reduction of the maximum photochemical efficiency of the photosystem II in *Citrus* leaves. Heber *et al.* (1996) showed that photorespiration is a useful physiological way to prevent photoinhibition under stomatal closure.

The dramatic decline of F_v/F_m and the rise in F_0 which was observed at temperatures above 47°C indicates the inhibition of the photosynthetic apparatus by heat stress (Bilger *et al.* 1987, Yamane *et al.*, 1997; Herppich *et al.*, 1994). However, as described for other mediterranean trees (Bilger *et al.*, 1987), serious heat stress can be observed above 45°C. In our study, any inhibition was reversible and F_v/F_m increased to predawn values as leaf temperature decreased during the afternoon. Short term experiments indicated that acclimatization to heat stress may occur. Weis & Berry (1988) showed for bean leaves that a 3-hour pre-adaptation at 40°C caused an increase in the tolerance limit by about 4°C. In potatoes a moderate increase of temperatures in 2.5 hours from 25°C to 38°C caused rapid photosynthetic acclimatization and increased the critical temperature from 38°C to 43°C (Havaux, 1993). Also, growth temperatures may largely influence temperature optimum photosynthesis (Björkman *et al.*, 1980). Under field conditions *Citrus* will be able to adapt to increasing temperature during the growing season. Desert evergreen species, which experience large seasonal changes in temperature show large changes in their thermal stability. The shift in thermal tolerance can be up to 10°C and even higher values have been reported (Berry & Björkman, 1980). Our experiments show for citrus that PSII acclimates to heat stress during the season. Thus, *Citrus grandis* seems to be well adapted to the high photon flux density and heat stress occasionally occurring in the Arava Valley.

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References

- Baker, N.R., 1993. Light-use efficiency and photoinhibition of photosynthesis in plant under environmental stress, In: Smith, J.A.C. & Griffiths, H. Water deficits, plant responses from cell to community, Environmental Plant Biology Series, βios Scientific Publisher, Oxford, 221-235
- Berry, J.A. & Björkmann, O., 1980. Photosynthetic response and adaptation to temperature in higher plants, *Ann. Rev. Plant Physiol.* 31: 491-543
- Bielorai, H., Levin, I. & Assaf, R., 1973. Irrigation of fruit trees, In: Yaron, B., Danfors, E., Y. Vaadia, Arid Zones Irrigation, Ecological Studies 5, Springer, Stuttgart, 397-404
- Bilger, W., Schreiber, U. & Lange, O.L., 1987. Chlorophyll fluorescence as an indicator of heat induced limitation of photosynthesis in *Arbutus unedo* L., In: Tenhunen *et al.* (eds.) Plant Response to Stress, 391-399
- Blanke, M. M., 2000. repair mechanisms following photoinhibition in Citrus. *Acta Horticulturae* No. 531 (this volume): 155-158
- Braake, M. & Allen, L.H., 1995. Gas exchange of Citrus seedlings at different temperatures, vapor-pressure deficits, and soil water contents, *J. Amer. Soc. Hort. Sci.* 120(3): 497-504
- Björkman, O., Badger, M.R. & Armond, P.A., 1980. Response and adaptation of photosynthesis to high temperatures, In: Turner, N.C. & Kramer, P.J. Adaptation of plants to water and high temperatures stress, Wiley-Interscience Publication, New York, 233-249
- Clifford, S.C., Kadzere, I. Jones, H.J. & Jackson, J.E., 1997. Field comparison of photosynthesis and leaf conductance in *Ziziphus maritiana* and other fruit tree species in Zimbabwe, *Trees* 11: 449-454
- Cohen, S., Mareshet, S., Guillou, L.L., Simon, J.-C. & Cohen, M., 1997. Response of citrus trees to modified radiation regimes in semi-arid conditions, *J. Exp.Bot.* 48: 35-44
- Hall, A.E., Camacho, B.S.E. & Kaufmann, M.R., 1975. Regulation of water loss by citrus leaves. *Physiologia Plantarum* 33: 62-65
- Havaux, M., 1993. Rapid photosynthetic adaptation to heat stress triggered in potato leaves by moderately elevated temperatures, *Plant Cell Environm.* 16: 461-467
- Heber, U., Bligny, R., Streb, P. & Douce, R., 1996. Photorespiration is essential for the protection of the photosynthetic apparatus of C3 plants against photoinactivation under sunlight, *Bot. Acta* 109(4): 307-315
- Herppich, M., Herppich, W.B. & von Willert, D.J., 1994. Influence of drought, rain and artificial irrigation on photosynthesis, gas exchange and water relations of the fynbos plant *Protea acaulos* (L.) Reich at the end of the dry season. *Bot. Acta* 107: 369-472.
- Herppich, W.B., Flach, B. M.-T., von Willert, D.J. & Herppich, M., 1997. Field investigations of photosynthetic activity, gas exchange and water potential at different ages in *Welwitschia mirabilis* during a severe drought, *Flora* 191: 59-66
- Herppich, W.B., 2000. Interactive effects of light and drought stress on photosynthetic activity and photoinhibition under (sub-) tropical conditions, *Acta Horticulturae* No. 531 this volume: 137-146
- Krall, J.P. & Edwards, G.E., 1992. Relationship between photosystem II activity and CO₂ fixation, *Physiologia Plantarum* 86: 180-187.
- Kriedemann, P.E. & Baars, H.D., 1981. Citrus Orchards, In: Kozlowski, T.T.: Water deficits and plant growth, Academic Press, 325-417
- Lange, O.L. & Meyer, A., 1979. Mittäglicher Stomatenschluß bei Aprikose (*Prunus armeniaca*) und Wein (*Vitis vinifera*) im Freiland trotz guter Bodenwasser-Versorgung, *Flora* 168: 511-528
- Larcher, W., Wagner, J., Neuner, G., Méndez, M., Jiménez, M.S. & Morales, D. 1991. Thermal limits of photosynthetic function and viability of leaves of *Persea indica* and *Persea americana*, *Acta Oecologia* 12 (4): 529-541
- Moreschet, S., Cohen, Y., Green, G.C., & Fuchs, M., 1990. The partitioning of hydraulic conductances within mature orange trees, *J. Exp. Bot.* 41: 833-839

- Singlair, T. R. & Allen, L.H., 1982. Carbon dioxide and water Vapour Exchange of Leaves on Field-Grown Citrus Trees, *J. Exp. Botany* 137: 1166-1175.
- Spiegel, R. & Goldschmidt, E.E., 1996: *Biology of Citrus, Biology of Horticultural Crops*, Cambridge University Press, 230 p.
- Veste, M. & Herppich, W.B., 1995. Influence of diurnal and seasonal fluctuations in atmospheric CO₂ concentration on the net CO₂ exchange of poplar trees, *Photosynthetica* 31(3): 371-378
- von Caemmerer, S. & Farquhar, G.D., 1981. Some relationships between the biochemistry of photosynthesis and the gas exchange of leaves, *Planta* 153: 76-387
- von Willert, D.J, Mattysek, R. & Herppich, W.B., 1995. *Experimentelle Pflanzenökologie, Grundlagen und Anwendungen*, Thieme Verlag, Stuttgart
- Yamane, Y., Kashino, Y. Koike, H. & Satoh, K., 1997. Increases in the fluorescence F₀ level and reversible inhibition of photosystem II reaction center by high-temperature treatment in higher plants., *Photosynth. Res.* 52: 57-64
- Werner, C., Correia, O. & Beyschlag, W., 1999. Two different strategies of mediterranean machia plants to avoid photoinhibitory damage by excessive radiation levels during summer drought, *Acta Oecologica* 20 (1): 119-123
- Weis, E. & Berry, J.A. 1988. Plants and high temperature stress, In: Long, S.F. & Woodward, F.I., eds., *Plants and Temperature*, Comp. Biol. Ltd., Cambridge, 329-346